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NOVEL DRUG TARGET**Field of the invention**

The present invention relates to a novel drug target. More precisely, glutathione transferase (GST) as target for treatment of cancer and other diseases responsive to inhibition of steroid hormone production. Preferably, the GST is GST A3-3.

Background of the invention

Prostate cancer and breast cancer are two major forms of malignant disease, which affect a significant proportion of the population. Tumor growth in both cases is often dependent on steroid hormones and an important therapeutic approach involves ablation of hormone production and blockage of the hormone receptor (1-3).

Steroid hormone biosynthesis proceeds from cholesterol to androgens (e.g. testosterone and dihydrotestosterone) and estrogens (e.g. progesterone and estradiol) via a series of metabolic intermediates. An obligatory step in each pathway leading to the respective hormones involves the isomerization of the Δ^5 -double bond to the Δ^4 -double-bond in the steroid structure. The isomerization is preceded by oxidation of the 3β -hydroxy compound into a 3β -keto steroid, catalyzed by 3β -hydroxysteroid dehydrogenase. This dehydrogenase has been shown to have an associated steroid isomerase activity.

Glutathione transferases, GSTs, occur in multiple forms (4) and are present in all cellular fractions. The mammalian GSTs can be divided into soluble and membrane-bound enzymes. They are traditionally regarded as detoxication enzymes that constitute the main cellular defense against electrophilic compounds that cause mutations, cancer and other degenerative diseases. However, it is also clear that some GSTs have specific roles in relation to physiologically relevant substrates. Therefore, the spectrum of GST activities is not limited to general detoxication of electrophiles, and some GSTs have roles in the metabolism of well-defined substrates. The recently discovered GST A3-3 appears to have such a role in double-bond isomerizations of steroids in hormone biosynthesis (5). The enzyme is present in steroidogenic organs such as testis, ovary, placenta and the adrenal gland, but not in significant amounts in other tissues such as liver, thymus, skeletal muscle and brain (5). A

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putative GST in the human adrenal cell line H293R is markedly induced by adrenocorticotrophic hormone (ACTH), a pituitary peptide that stimulates steroid hormone synthesis (6).

Furthermore, it is known that GSTs are inhibited by a wide variety of agents *in vitro* (4), but inhibition data have not previously been obtained for the recently discovered GST A3-3 (5). Some GST inhibitors have been shown to be effective in cellular systems and in clinical trials (7-9).

Summary of the invention

According to the present invention glutathione transferase (GST), preferably GST A3-3, is provided as a new target for chemotherapy, based on its contribution to double-bond isomerizations in steroid biosynthesis. GST A3-3 has selective tissue distribution and shows high catalytic activity in the isomerization of both Δ^5 -androstene-3,17-dione and Δ^5 -pregnen-3,20-dione (Fig. 1). The present inventor has shown that the catalytic efficiency of GST A3-3 is 200-fold higher than the steroid isomerase activity of 3 β -hydroxysteroid dehydrogenase. The invention is primarily concerned with cancer in the prostate, but the principle of inhibiting steroid hormone production is also applicable to steroid-responsive cancer in the breast and in other organs. Further, it is applicable to other steroid hormone-dependent diseases such as Cushing's syndrome.

Thus, in a first aspect the invention relates to the use of glutathione transferase (GST) as a drug target for screening of compounds that inhibit the activity of GST for treatment of steroid hormone dependent diseases, such as for treatment of cancer, preferably prostate cancer and breast cancer.

The GST is preferably GST A3-3. Preferably, pharmaceutically acceptable compounds, which inhibit the activity of GST A3-3, are screened for. Thus, the present invention relates to a method for screening of compounds or drug candidates that modulate, preferably inhibit, GST in which method GST is used as a drug target. Such a screening assay may for example be performed as in high throughput screening.

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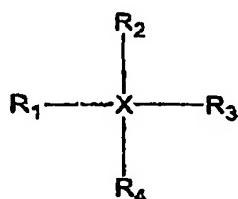
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In a second aspect, the invention relates to the use of inhibitors against GST A3-3 for production of a drug for treatment of steroid hormone dependent diseases, such as for treatment of cancer, preferably the cancer is prostate cancer or breast cancer.

Examples of compounds to be used according to the invention include GST inhibitors having the following formula:



wherein R₁, R₂, R₃ and R₄ can be alkyl groups, such as methyl, ethyl, propyl, butyl, pentyl, hexyl; aryl groups, such as phenyl or substituted phenyl, preferably substituted with lower alkyl, hydroxyl or alkoxy groups; or chemical derivatives or combinations of these groups; the R₁, R₂, R₃ and R₄ groups can be linear; branched, such as substituted with lower alkyl, hydroxyl or alkoxy groups; or cyclic, such as cyclopentyl and cyclohexyl; the R₁, R₂, R₃ and R₄ groups can contain heteroatoms such as O, S, and N. The inhibitors can be stereoisomers depending on the nature and spatial orientation of the groups surrounding X; two, three or four of the R₁, R₂, R₃ and R₄ groups can be linked together and have a bidentate, tridentate or tetradeionate coordination with the central atom X; Alternatively, one, two, three or four of R₁, R₂, R₃ and R₄ can be Cl, Br, I, O, S, Se, carboxylate ions such as acetate and homologs, or other chemical ligands with an electron-donating group coordinated to X.
 X= Ge, Sn, Pb or similar electrophilic atoms.

The GST inhibitors preferably contain tin (Sn) as electrophilic atom, since such compounds combine moderate toxicity (10,11) with strong inhibition of the target enzyme. The tin atoms of the inhibitors can have different oxidation states, such as Sn(II) or Sn(IV), and the coordination number of the ligands can be 2, 3, 4, 5 or 6.

Preferably, one of R₁-R₄ is Cl, Br or acetate and the other substituents are ethyl, butyl or phenyl.

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In a third aspect, the invention relates to a method for treating cancer or steroid hormone dependent diseases, comprising administering a non-steroidal compound that inhibits the enzymatic activity of GST A3-3 to a human in need of such a treatment.

The human is in need of cancer treatment or treatment of other steroid hormone dependent diseases, such as Cushing's syndrome.

In one embodiment the human is a male who suffers from prostate cancer. In another embodiment the human is a female who suffers from breast cancer.

Detailed description of the invention

The invention will be described more closely below in association with some non-limiting examples and figures.

Fig. 1. Metabolic pathways leading from cholesterol to steroid hormones such as testosterone (and further to dihydrotestosterone) and progesterone (and further to estradiol). The hormones act via binding to the androgen and estrogen receptors, respectively, and promote growth of hormone responsive prostate and breast cancer. GST A3-3 catalyzes essential steroid isomerizations in the respective pathways and the invention involves this enzyme as a target for hormone responsive disease.

Fig. 2. Alternative reactions for measuring the inhibition of GST A3-3 in vitro. All three reactions can be monitored spectrophotometrically using purified enzyme and glutathione (GSH): (A) Δ^5 -androstene-3,17-dione; (B) 1-chloro-2,4-dinitrobenzene; and (C) phenethylisothiocyanate. Addition of an inhibitor will decrease the rate of the reaction catalyzed by GST A3-3.

EXPERIMENTAL PROCEDURES

Materials—1-Chloro-2,4-dinitrobenzene (CDNB) and reduced glutathione (GSH) can be purchased from Sigma (St. Louis, MO), phenethylisothiocyanate from Aldrich (Milwaukee, WI), and Δ^5 -androstene-3,17-dione from Steraloids Inc. (Newport, RI).

Expression and purification of GSTs—Human GST A3-3 and its homologous GST proteins of the Alpha class were expressed from corresponding cDNA carried by the pET-21a(+) vector in *E. coli* BL-21(DH3) (5). The cells were grown to OD₆₀₀=0.7 and expression was induced by

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addition of 1 mM IPTG. The cells were grown for four hours, collected by centrifugation, and lysed using ultrasonication. The lysate was desalted on a PD-10 gel filtration column (Amersham Biosciences) and the proteins were eluted in 20 mM sodium phosphate, pH 7.0, and were subsequently loaded onto a HiTrap SP cation exchanger (Amersham Biosciences). The proteins were eluted using a salt gradient. This single purification step yielded highly pure enzymes as confirmed by SDS-PAGE stained with Coomassie Brilliant Blue.

Specific activity measurements—The specific activities of GST A3-3 were determined for the isomerization reaction with Δ^5 -AD (Fig. 2A), the conjugation reaction with 1-chloro-2,4-dinitrobenzene (CDNB) and GSII (Fig. 2B), and for the addition of GSII to phenethylisothiocyanate (Fig. 2C). The reactions were monitored spectrophotometrically at 30 °C. The isomerization of 100 μ M Δ^5 -AD was followed at 248 nm in 25 mM sodium phosphate buffer, pH 8.0, in the presence of 1 mM GSH. The extinction coefficient for the product Δ^4 -AD is 16,300 M⁻¹cm⁻¹ (12). Specific activity measurements were performed in 0.1 M sodium phosphate, pH 6.5, with 1 mM CDNB in the presence of 1 mM GSII as described (13), and with 0.1 mM phenethylisothiocyanate in the presence of 1 mM GSII (5).

Examples of specific inhibitors of Alpha class GSTs

Enzyme activities were determined in the standard assay system and the concentration of the inhibitor giving 50 % inhibition of the activity (IC_{50}) was determined.

Even if the compounds are inhibiting several GSTs, some inhibitors display high selectivity for a given GST (4). The present inventor has shown that this applies also to homologous members of the same GST class (Table 1). Selective inhibition is desirable to avoid interference with non-targeted GST-catalyzed reactions and to minimize toxic side effects. Without any extensive screening, inhibitors of GST A3-3 effective in the nanomolar concentration range have already been identified. These inhibitors also display selectivity among GST A3-3 and other human Alpha class members (Table 1). However, the related GST A1-1 has approximately 5% of the specific activity of GST A3-3 in the isomerization of androstenedione (5,12), and it may be advantageous to inhibit GST A1-1 in addition to GST A3-3. By use of multivariate cluster analysis of inhibition data it is possible to optimize discrimination among the enzymes.

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Table 1. Differential inhibition of Alpha class glutathione transferases demonstrated by using organometallic compounds. The IC₅₀ values are the inhibitor concentrations giving 50% inhibition of the GST-catalyzed reaction.

Inhibitor	IC ₅₀ Values (μM)			
	GST	A1-1	A2-2	A3-3
Et ₃ GeCl	56	0.8	67	
Bu ₃ SnAc	0.018	0.41	0.018	
Et ₃ SnBr	5.7	0.19	0.69	
Ph ₃ PbCl	0.0046	0.084	0.013	
Ph ₃ SnAc	0.16	Nd	0.16	
Et ₃ PbCl	2	Nd	2.3	
Ph ₃ PbBr	0.0086	Nd	0.16	

Et, Bu, and Ph are ethyl, n-butyl, and phenyl, respectively; Nd = not determined.

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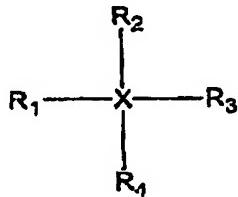
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CLAIMS

1. Use of glutathione transferase (GST) as a drug target for screening of compounds that inhibit the activity of GST for treatment of steroid hormone dependent diseases.
2. Use according to claim 1, for treatment of cancer.
3. Use according to claims 1 or 2, wherein GST is GST A3-3.
4. Use of inhibitors against GST A3-3 for production of a drug for treatment of steroid hormone dependent diseases.
5. Use according to claim 4 for production of a drug for treatment of cancer.
6. Use according to claim 5, wherein the cancer is prostate cancer.
7. Use according to claim 5, wherein the cancer is breast cancer.
8. Use according to any of the claims 4-7, wherein the inhibitor is a compound having the following formula:



wherein R₁, R₂, R₃ and R₄ can be alkyl groups, such as methyl, ethyl, propyl, butyl, pentyl, hexyl; aryl groups, such as phenyl or substituted phenyl, preferably substituted with lower alkyl, hydroxyl or alkoxy groups; or chemical derivatives or combinations of these groups; the R₁, R₂, R₃ and R₄ groups can be linear, branched, such as substituted with lower alkyl, hydroxyl or alkoxy groups; or cyclic, such as cyclopentyl and cyclohexyl; the R₁, R₂, R₃ and

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R₁ groups can contain heteroatoms such as O, S, and N; alternatively, one, two, three or four of R₁, R₂, R₃ and R₄ can be Cl, Br, I, O, S, Se, carboxylate ions such as acetate and homologs, or other chemical ligands with an electron-donating group coordinated to X;
X= Ge, Sn, Pb or similar electrophilic atoms;

as well as stereoisomers of the inhibitor.

9. Use according to claim 8, wherein X is Sn.

10. Use according to claim 8 or 9, wherein one of R₁- R₄ is Cl, Br or acetate and the other substituents are ethyl, butyl or phenyl.

11. Use according to any one of the claims 4-10, wherein the inhibitor also inhibits GST A1-1.

12. A method for treating cancer or steroid hormone dependent diseases, comprising administering a non-steroidal compound that inhibits the enzymatic activity of GST A3-3 to a human in need of such a treatment.

13. A method according to claim 12, wherein the human is a male who suffers from prostate cancer.

14. A method according to claim 12, wherein the human is a female who suffers from breast cancer.

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Abstract

The present invention relates to a novel drug target, glutathione transcrasc (GST), preferably GST A3-3, as target for treatment of cancer and other diseases responsive to inhibition of steroid hormone production. The present invention also relates to a method for screening of compounds or drug candidates that modulate, preferably inhibit, GST, in which method GST is used as a drug target. The invention further relates to the use of inhibitors against GST A3-3 for production of a drug for treatment of steroid hormone dependent diseases, such as for treatment of cancer, preferably prostate cancer or breast cancer. The present invention also relates to a method for treating cancer or steroid hormone dependent diseases, comprising administering a non-steroidal compound that inhibits the enzymatic activity of GST A3-3 to a human in need of such a treatment.

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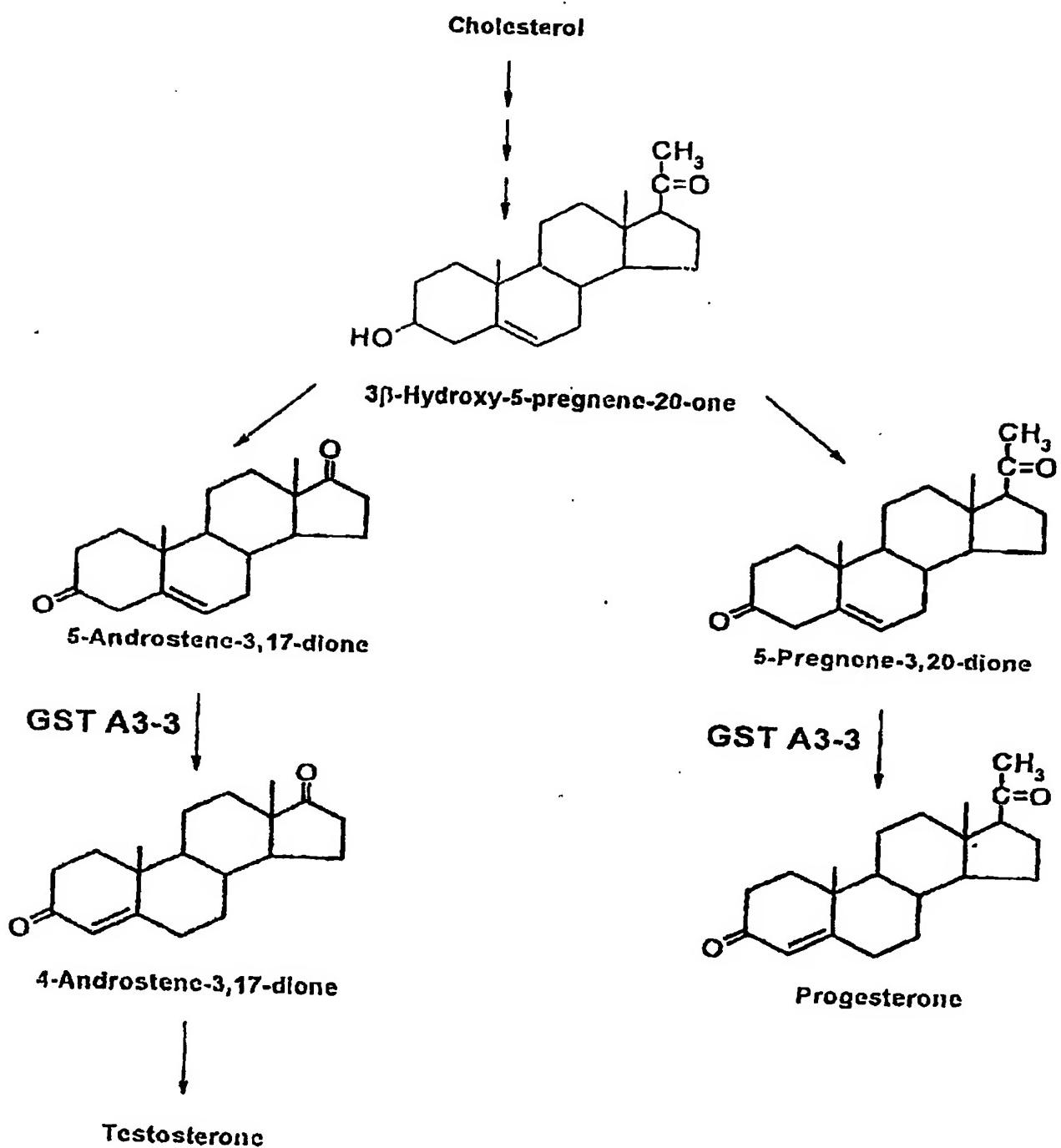
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FIG. 1

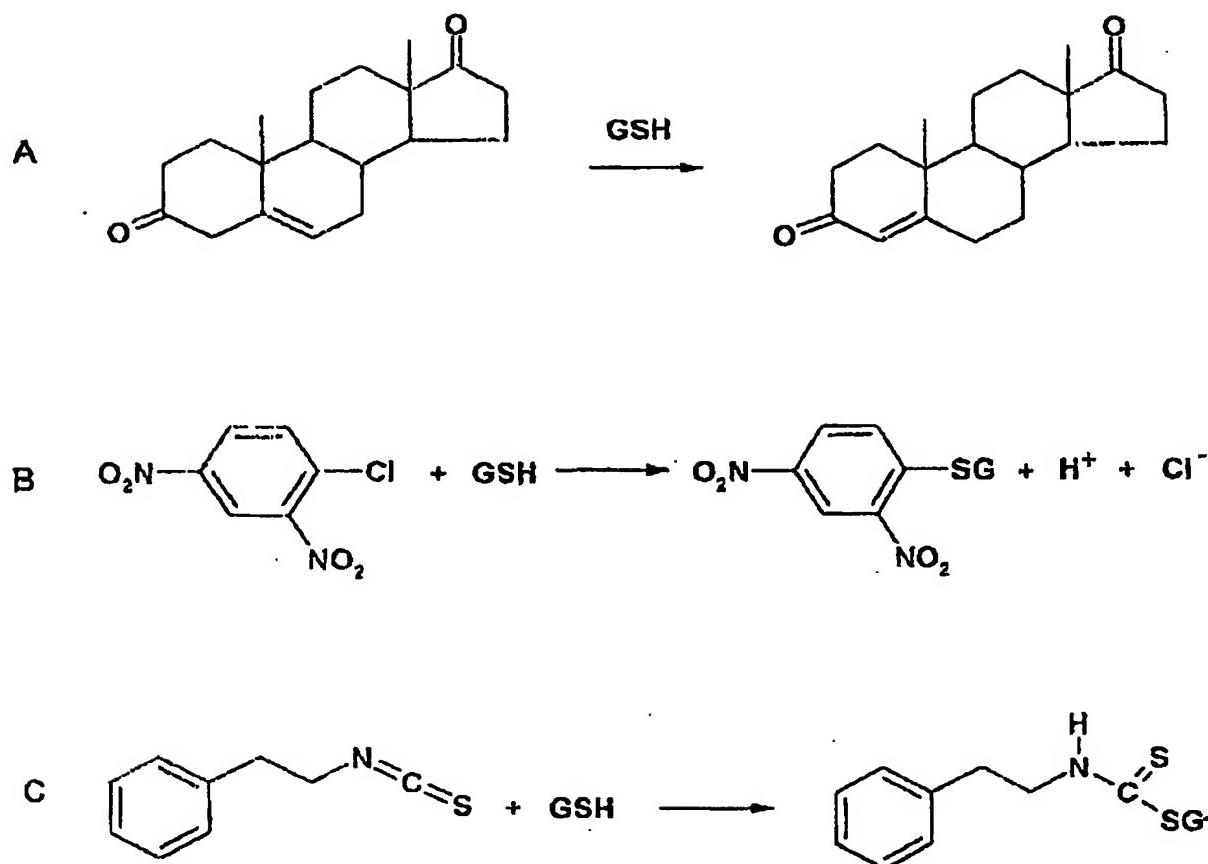


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FIG. 2



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